

What is Lymphology – Prospects in Human Studies*

Waldemar L. Olszewski

Dept. of Surgical Research and Transplantology, Medical Research Center, Polish Academy of Sciences, Warsaw, Poland, and Lab. of Hematology and Lymphology, Norsk Hydro's Institute for Cancer Research, The Norwegian Radium Hospital, Oslo, Norway

If we asked a medical graduate what is the lymphatic system, the answer would be: The lymphatic system is an anatomical structure composed of channels, with the principal function to maintain the blood volume by returning fluid and protein molecules which leak from the blood capillaries to the interstitial space, to the general circulation. In addition, there are circulating lymphocytes and lymphoid organs playing an important role in the process of defense against infection and tumor growth.

Using this commonly accepted approach our image of the lymphatic system is limited to its structural elements like cells, vessels, organs and to the function of these elements within the system, e.g. lymph protein transport, cooperation of various lymph cell populations in the lymph node, etc. However, thinking teleologically the mission of the lymphatic system should certainly be more wide-reaching. The lymphatic system serves the whole living organism in the process of homeostasis. In what way? By maintaining a physiological environment of each individual cell in the non-lymphoid tissues of the body. This microenvironment includes the fluid and ground substance surrounding the cell, as well as the neighbouring cells with

their genetically defined self. One could, then, define lymphology as the biomedical discipline dealing with *problems of regulation of the cellular microenvironment*. Then, the principal tasks of the lymphatic system are:

- a) maintaining of a most favorable composition of the mobile intercellular fluid and the ground substance for the tissue cells, their integrity and function,
- b) transporting away and processing chemical products released from cells, as well as their subcellular shedded structures, e.g. membrane receptors,
- c) eliminating of dying or mutant cells,
- d) removing of non-self organic (e.g. bacteria, viruses) and inorganic (e.g. carbon, silica) particles entering the intercellular space from environment.

A. This is the individual cell in a living context of the neighbouring cells and the surrounding, amorphous ground substance which is source of signals setting the lymphatic system into motion. Let us look at the lymphatic system as subordinate to the tissue non-lymphoid cells and standing under their orders. This approach has many important implications. First of all it allows to study the function of the lymphatic system in the context of the living non-lymphoid cells and not as a system working for itself. Here are some examples:

- a) Studying the process of regulation of the interstitial volume we take into consideration the physical factors responsible for the cell

* Excerpts from the lecture "New Horizons in Lymphology" at the closing session of the Congress of the International Society of Lymphology, Montreal, Canada, September 25, 1981

lary and lymphatic transport of water and macromolecules, but disregard the possible role of the nonlymphoid cells bathing in the interstitial fluid in the regulation of its volume and composition. Beyond doubt, cells themselves can mediate chemically the capillary permeability, regulate their water uptake from the interstitial fluid, control the chemical composition of the ground substance. The regulation of the interstitial volume cannot be a one-way, *vessels-intercellular space-cells*, process, without any feedback signals from the cells.

b) There is a continuous process of leukocyte migration through the non-lymphoid tissues. These cells presumably play a principal role in recognizing and removing exogenous and mutant-self antigens and transmitting information about their appearance in the tissue to the higher regulating centers like lymph nodes, spleen, bone marrow. The signals for the lymphocytes to leave blood circulation and enter the tissue space originate from the parenchymatous cells or their chemical products. Thus, the regulation of lymphocyte and monocyte traffic across the non-lymphoid cells is dictated primarily by the tissue events.

c) Does the concentration of the free moving proteins of different molecular weight and their biological activity in the interstitial space depend only on the capacity of the capillary and lymphatic transport or can the parenchymatous cells also influence protein concentration around them? Over billions of years of evolution cells have been organizing their environment. Today, there is a different quantitative composition of free-moving proteins in the interstitial space compared to the blood. It is likely that cells can to a large extent control the local protein concentration not only by excreting their proteinaceous products but also by increasing influx from blood circulation by enhancing capillary permeability, e.g. in an enhanced metabolic state. These examples have been presented to substantiate the rightness of reasoning that the functions of the lymphatic system are regulated primarily by signals originating from the parenchymatous cells of the non-lymphoid tissues.

B. There is another problem concerning the understanding of the function of the lymphatic system. The anatomical territory of the lymphatic system comprises the interstitial space, lymph vessels, organized lymphoid tissue and its locomotive messengers- the migratory cells. All of these elements are functionally interrelated. Dissociation of the lymphatic transport pathways from the lymphoid tissue, while analyzing the function of the lymphatic system, is no longer tenable.

Two examples support this notion. One, the lymphatic endothelial cells line the channels serving fluid transport, but at the same time possess the phagocytic activity, a function ascribed to the immune system. Another, the capacity of fluid transport of lymph vessels restricts the kinetics of antigen transport away from the interstitial space and its delivery to the sites of elimination – lymph nodes.

C. The lymphatic system is one of the elements of the entire organization of body homeostasis. It can not work independently of the nervous and endocrine system. It has been documented recently, that the peripheral, as well as the central nervous system, play a distinct but still largely undefined role in the immune regulation. However, the operation of afferent and efferent pathways to and from the neuroendocrine structures has thus far not received any serious consideration in relation to the function of the lymphatic system. There are indications that the primary immune response of rats to a particular antigen, SRBC or soluble antigens is followed by several-fold increase in corticosterone levels and also temporal changes in thyroxine levels. An evident increase in firing rates of neurones of ventromedial nuclei in the rat hypothalamus after intraperitoneal injection of SRBC was observed. Presumably, this neuroendocrine mechanism functions when a critical threshold of lymphoid tissue activation is reached, sufficient to elaborate products serving as a signal to the hypothalamus. Further knowledge of the profile of hormonal responses and the neuroendocrine-lymphoid tissue and lymph transport pathways interrelation will be required.

Summarizing what was said above, the lymphatic system is an organization: a) composed of the functionally interrelated lymphoid tissue and transportation pathways, b) operating for maintaining of a proper environment of the non-lymphoid cells in the organized tissues and securing their genetically restricted self, especially those tissues having contact with the outer world like skin, gut and lungs, and c) integrated with the nervous and endocrine systems.

Which are the main problems we should work on in order extend our knowledge of the *in vivo* function of the lymphatic system?

These are:

- 1) maintaining of an appropriate chemical and physical environment for cells in the non-lymphoid tissues with special emphasis on the regulatory signals released from these cells,
- 2) function of the immune cells migrating through the non-lymphoid tissues,
- 3) cooperation of the immune cells at various regulatory levels *in vivo* (lymph nodes, spleen, bone marrow, etc.),
- 4) integration of the regulatory processes of the lymphatic, nervous and endocrine systems,
- 5) more emphasis on studies in humans with the non-invasive techniques.

In order to go into more details and ask more specific questions I show a trivial picture illustrating the functional anatomy of the lymphatic system (Fig. 1). Several physiological questions may be asked with regard to the events in its various anatomical regions. The entire system has been divided into four interrelated functional areas. The basic one named "community" illustrates a normal tissue with a blood capillary and an initial lymphatic and a group of parenchymatous cells. The cells lead an intensive life restricted by supplies and transport away of waste products and controlled by immigrant immune cells. There are proteins of various molecular weight in plasma and interstitial fluid presented as suitcases and bags and mononuclear cells and some polymorphs also carrying on their surface specific proteins. The protein molecules are transported through the capillary membrane which acts as a molecular sieve. What is the biological activity of these proteins in the interstitial space which is, no doubt, influenced both quantitatively and qualitatively by the restricted transport across the capillary membrane? How does the capillary membrane and the ground substance organization affect the transport of antibiotics and chemotherapeutics to the tissue fluid? Which are the subcellular organelles and membrane fragments from the disintegrated

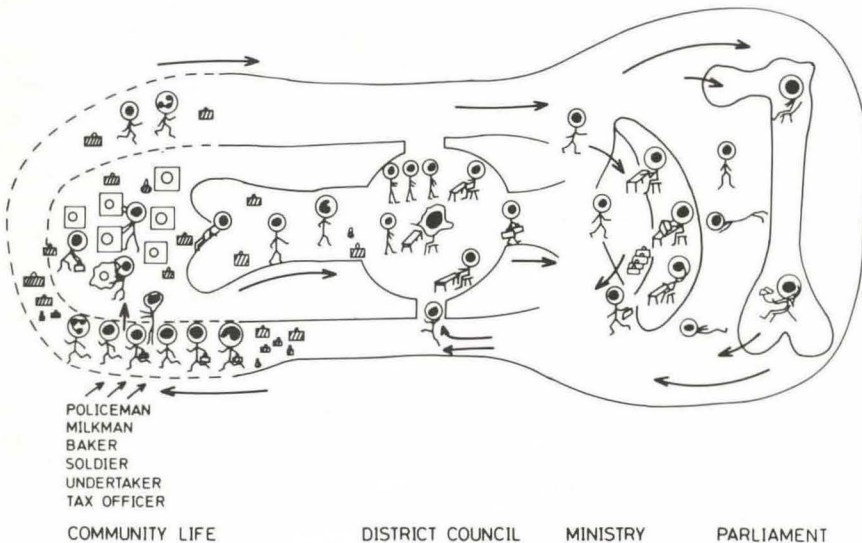


Fig. 1

ed dead cells released into the tissue fluid and lymph? Do these cellular elements, as well as the chemical substances released from cells act as signals informing the higher regulatory centers via lymph stream about the tissue integrity?

There are leukocytes in blood capillaries which look morphologically much alike. However, they are at various stages of maturity and possess different functional capacities. How could we characterize them functionally? Which cells leave blood circulation heading for the interstitial space and initial lymphatics? What is their function? Do they eliminate or limit spread of neoplastic cells? What are the forces moving the free interstitial fluid into initial lymphatics and further along the prenodal lymph vessels. The speed of lymph flow does not only mean the speed of fluid movement, it is also the quickness of transmitting of immune information with shedded antigen, primed phagocytes and lymphocytes from the tissue to the regional lymph node and other lymphoid organs.

Proteins and cells travel along lymphatics to the lymph nodes (district council), and once they reached blood circulation, also to the spleen (ministry) and bone marrow (parliament) (Fig. 1). One of the principal events occurring in the lymphatic organs is communication between lymphocytes. The cell communication events in the immune system is to control the correct antigen specificity and the appropriateness of the response. What do we know about the kinetics of cell communication in the lymph nodes, spleen and bone marrow? Is the spleen the principal site for elimination of particles and cells with non-self antigens or altered surfaces or rather the institution educating lymphocytes and monocytes? What is the role of the liver in lymphocyte recirculation? Does this organ only eliminate damaged cells or are there also subsets specifically migrating to the liver? Why do the lymphocytes assembly in shock or after steroid administration in the bone marrow?

We approached some of these problems in our human studies. The excerpts from our works will be presented.

Studies in humans on the function of the lymphatic system give us the most useful practical informations. The insight into the interstitial space and prenodal lymph in man can be obtained by drainage of the superficial lymphatics of the calf, the technique worked out by A. Engeset et al. (1973). This technique has undergone, in our hands, some refinements so that now we can obtain 10–20 ml of lymph from one vessel per day (Olszewski 1977a). The method of a continuous drainage of lymph from the limb has enabled the studies of the influence of physical factors like gravity, temperature and venous congestion and of local immune stimulation on lymph flow, pressures, chemical and cellular composition.

A. Immune and other biologically active proteins in the interstitial fluid and lymph. The concentration of proteins in serum is relatively stable. Also, the proportions of proteins of various molecular weight remain in serum rather constant. In the interstitial fluid and lymph the situation is different. Both, the total protein concentration and concentrations of single proteins undergo fluctuations dependent on the changing conditions of the capillary filtration, volume of the interstitial space, lymphatic transport away of tissue fluid, etc. These, in turn, depend on the capillary permeability coefficient, filtration pressures, filtration surface area, changes of interstitial fluid pressures generated by muscular contractions, contractility force of lymphatics, etc. Thus, the protein concentration in the mobile interstitial fluid surrounding cells is not only significantly lower than in serum but also the levels of individual proteins are inversely proportional to their molecular weight (Engeset et al. 1977a, Olszewski et al. 1977b, 1977c). E.g. the average level of IgG in lymph obtained from human leg skin is 15–20%, of IgM 6–10% of that of serum. There might be also a difference in immune protein activity in tissue fluid and lymph compared with serum as for example with the C3 component (Olszewski et al. 1977d, 1978). The concentration and activity of complement inhibitors in tissue fluid is higher than of the components they are acting upon (Olszewski et al. 1982a). Gravitational forces influence directly

transcapillary protein transport in the most dependent parts of the body. However, we have found that the regulatory mechanisms of microcirculation are extremely efficient and the change from the lying to the upright position does not increase the capillary permeability for large molecular weight proteins (Olszewski et al. 1977c, 1979).

In order to properly understand the events developing in the tissue space we should be constantly aware of the levels of proteins and other substances in the interstitial fluid. However, the estimation of an actual concentration of protein in the interstitial fluid based on their measurement in lymph is rather complex. This is due to the time lag necessary for the tissue fluid to reach the drained lymph vessel and collecting cannula. This in turn is dependent on the rate of tissue fluid formation, tissue compliance — stress relaxation, and forces promoting tissue fluid and lymph flow. Engeset et al. (1979) measured in the human leg the time period necessary for the i.v. injected labelled protein to appear in lymph and the time of equilibration of labelled albumin between serum and lymph. The first appearance of labelled protein in sampled lymph was observed in less than 2 h. The equilibrium between serum and lymph was reached after 26 h. The patients were studied during their normal daily activities and night rest. Had we studied them lying in bed for several days, the time for equilibration would have probably been even longer. The results of this study have evident implications for understanding of the kinetics of extravascular distribution of drugs and substances bound to proteins.

B. Antibiotic penetration into tissue fluid and lymph. Prenodal lymph may be an extremely useful source of information on tissue fluid concentration of antibiotics and chemotherapeutics. Monitoring of antibiotic concentration in serum does not give reliable information on the level of drug in foci of infection. The widely accepted use of the concentration obtainable in serum by standard dosage of an antimicrobial agent as a guideline for setting of breakpoints in sensi-

tivity and resistance of a pathogen to a drug may be unreliable. In the studies of Bergan et al. (1979) ampicillin showed a delayed appearance, lower peak concentrations and protracted decay in lymph as compared with serum. Thus, the breakpoint between sensitivity and resistance of bacteria to antimicrobial agents should be set at a much higher peak concentration than that obtainable in serum.

C. Cells in human prenodal lymph.

As mentioned before human prenodal lymph contains lymphocytes, macrophages and erythrocytes (Engeset et al. 1977b). Which are the leucocyte populations which leave blood circulation and enter the tissue space in the non-lymphoid tissues? What is their function? Can they control cancer cell metastases?

Lymphocytes migrate actively across the non-lymphoid tissues. The cell movement is accomplished by three closely correlated mechanisms: generation of driving force, adaptation of cell shape, and formation of attachment sites. There are changes in direction of locomotion of locomotive cells, short tracks and reduction of speed imposed by the texture of their environment. But the main question remains in what tissue compartment the cells actually locomote. There are no empty spaces and what appears as such contains a network of proteoglycans associated with collagen. Propulsion by mere physical pressure generated by locomoting cells sufficient, or lysis of host constituents is necessary. The tempo of migration of lymphocytes through the tissue to lymphatics is different from that of red cells. There is a relatively high concentration of lymphocytes in lymph when the lymph flow is low, quite opposite to red blood cells. This indicates that lymphocytes have their own tempo of active migration, and as we know from more detailed studies is relatively constant throughout the 24 hrs (Engeset et al. 1977b).

There appears to be some selection of migrating cells at the level of the capillary bed, which certain cell types are selected to enter the tissue space and lymphatics (Engeset et al. 1974, 1977c, Godal and Engeset 1978). Cell populations in afferent lymph of human

leg were defined by surface characteristics and cytotoxic activity in normal men (Lukomska et al. 1981). A higher percentage of E-rosette forming cells was found in lymph (78.5) than in blood (60.0). The percentage of lymph EA-RFC was 10.3 of EAC-RFC 13.1 and of surface immuno-globulin carrying cells 3.0. In blood 20.6 percent of cells formed EA-rosettes, 23.0 EAC-rosettes, 5 contained surface immunoglobulins. The differences between lymph and blood EA- and EAC-RFC were statistically significant ($p < 0.05$). The natural cytotoxicity against K 562 cells was 6 times lower in lymph as compared to blood ($p < 0.05$). The study indicates that B-cells have a limited tendency toward leaving the blood circulation and migrating through tissues. Moreover natural killer cells do not seem to belong to the recirculating pool of lymphocytes.

We made also use of the monoclonal antibodies, characterizing the cell populations which migrate into the normal human skin and which having traversed the tissue, could be recovered from the afferent lymph vessels (Olszewski et al. 1982b). Significant differences were apparent between the types and proportions of cell populations in lymph and blood. The percentage of OKM1⁺ cells (monocytes, null cells) in lymph was low when compared to that of blood. It may be noted that the OKM1 antibody did label only about 40 percent of the large mononuclear macrophage-like lymph cells supposed to be Langerhans cells. The percentage of OKT3⁺ (T cells) in lymph was higher than in blood as was that of the OKT4⁺ (inducer/helper) subset, while cells of the OKT8⁺ (suppressor/cytotoxic) subset were found to be more numerous in blood. The OKI a1⁺ cell population consisted of large veiled mononuclear cells and only very few small cells. The former were not detected in blood. Surprisingly, the large mononuclear cells present in lymph reacted with OKT6 antibody specific for cortical thymocytes. The discovery of high proportions of T cells, cells bearing Ia-like antigens, and a high inducer/suppressor ratio in normal human prenodal lymph reflects the intensity of "physiological" immune processes in the skin.

In another project we investigated the responsiveness to polyclonal mitogens of cell populations which could be recovered from the afferent lymph vessels (Olszewski et al. 1982c). A high spontaneous transformation rate of lymph mononuclear cells was observed after they have been shortly cultured *in vitro*. The ³H-TdR incorporation was after a 24 h culture 2-times, and after a 72 h culture 5-times higher than by the PBM cells of the same subject. The lymph cells responded actively to very low concentrations of PHA, ConA and PWM. These concentrations were too low to activate the PBM cells. The mitogen concentration response curves of lymph cells were significantly higher and reaching peaks at other concentrations than of PBM cells. The population revealing the high auto-transformation rate was found to belong to the OKT4⁺ – enriched subset (inducer/helper). This subset had also the highest rate of incorporation of ³H-TdR after stimulation with PHA. The findings of a high spontaneous activation of lymphocytes which trafficked through the skin and classification of this highly reactive subset as an OKT4 positive and highly responsive to mitogens, reflect the *in vivo* immune events in the normal skin. The highly reactive subset may belong to the autoreactive cells active in the autologous MLR.

Another problem of our interest in the studies on the physiology of the human lymphatic system has been the active transport of lymph in collecting vessels. We have found (Olszewski et al. 1968; Olszewski and Engeset, 1979a, b) that intrinsic contractions of human leg lymphatics generate pressure necessary for propelling lymph.

To study the efficiency of intrinsic contractions of lymphatics in propelling lymph in human legs, the end and lateral pressures and lymph flow were measured in leg subcutaneous lymph vessels in 25 normal men in a horizontal and upright position, during rest and during contractions of the foot and calf muscles (Olszewski and Engeset 1980). Systolic lymph end pressures generated by intrinsic contractions of lymphatics ranged between 12 and 70 mmHg, but in some cases reached values above 100 mmHg. Systolic lymph lateral pressures with free lymph flow were lower and

ranged between 5 and 30 mmHg ($p < 0.01$). There was almost no hydrostatic component of lymph pressure. Contractions of foot and calf muscles in the horizontal as well as in the upright position did not significantly effect systolic end pressures, but slightly increased lateral pressures ($p < 0.01$). There was an increase in the frequency of pulse waves ($p < 0.01$), but pulse amplitudes did not change. Lymph flow occurred in the resting human limbs and during limb muscle contractions only during the lymphatic pulse waves. There was no flow in the periods between the waves. However, the mean lymph flow was higher during muscular contractions than during the resting period ($p < 0.01$). External massaging of the foot did not produce any rise in lymph pressure, but the frequency of pulse waves and lymph flow increase. Filling of the lymphatics with lymph by external massaging or injection of fluid evoked intrinsic contractions. The pressure threshold for evoking contractions varied in different subjects from 5 to 25 mmHg. We have drawn the conclusion that intrinsic contractions of lymphatic collectors in the human leg is the main factor responsible for lymph flow during rest of the extremity.

I have presented excerpt from some works on the physiology of human lymphatic system, not mentioning any of the fascinating studies on the radiology, surgery and pathology or experimental investigations in animals carried out by the participants of our convention. This has been done on purpose. Whereas a spectacular progress has been made in the radiological diagnostic procedures, radiotherapy, *in vitro* pathophysiological studies and development of surgical techniques, our knowledge of the *in vivo* function of the lymphatic system in a normal man remains still very limited. More efforts in this direction are necessary.

Acknowledgements

The author wants to thank Dr. A. Engeset for the cooperation, discussions and continuous support which made the Polish-Norwegian joint-studies possible.

The studies quoted in this article were supported by a grant from the Norwegian Cancer Society and the Polish Academy of Sciences 10.5. The generous

help of the ASTRA LÄKEMEDEL, Södertälje, Sweden is acknowledged.

References

- Bergan, T., A. Engeset, W.L. Olszewski, R. Solberg: Pharmacokinetics of bacampicillin and mecillinam in plasma and peripheral lymph. *Lymphology* 12 (1979) 85
- Engeset, A., B. Hagar, A. Nesheim, A. Kolbenstvedt: Studies of human peripheral lymph. *Lymphology* 6 (1973) 1
- Engeset, A., S.S. Froland, K' Bremer: Studies of human peripheral lymph in patients with chronic lymphocytic leukemia. *Scand. J. Hematol.* 13 (1974) 93
- Engeset, A., W.L. Olszewski, P.M. Jaeger, J. Sokolowski, L. Theodorsen: Twenty four hour variation in flow and composition of leg lymph in normal men. *Acta Physiol. Scand.* 99 (1977a) 140
- Engeset, A., J. Sokolowski, W.L. Olszewski: Variation in output of leucocytes and erythrocytes in human peripheral lymph during rest and activity. *Lymphology* 10 (1977b) 198
- Engeset, A., K. Bremer, S.S. Froland: The cellularity of peripheral lymph in lymphomas. *Prog. Lymphol.* Ed. R.C. Mayall and M.H. Witte, Plenum Press, New York 1977c, 245-250
- Engeset, A., M. Aas, W.L. Olszewski, J. Sokolowski: Time of exchange of ^{131}I -labelled albumin between plasma and peripheral lymph in man. *Lymphology* 12 (1979) 77
- Godal, T., A. Engeset: A preliminary note on composition of lymphocytes in human peripheral lymph. *Lymphology* 11 (1978) 208-210
- Lukomska, B., W.L. Olszewski, A. Engeset: Immunologic characteristics of human peripheral lymph cell populations. *Lymphology* 13 (1980) 186
- Olszewski, W.L., S. Kruszewski, L. Zgliczynska et al.: Observations on movements of lymphatics in patients with lymphedema of limbs. *Pr. Tyg. Lek.* 23 (1968) 1345
- Olszewski, E.L.: Collection and physiological measurements of peripheral lymph and interstitial in man. *Lymphology* 10 (1977a) 137
- Olszewski, W.L., A. Engeset, P.M. Jaeger, J. Sokolowski, L. Theodorsen: Flow and composition of leg lymph in normal men during venous muscular activity and local hyperthermia. *Acta Physiol. Scand.* 99 (1977b) 149
- Olszewski, W.L., A. Engeset, J. Sokolowski: Lymph flow and protein in the normal male leg during walking, lying and getting up. *Lymphology* 10 (1977c) 178
- Olszewski, W.L., A. Engeset, H. Lukasiwicz: Immunoglobulins, complement and lysozyme in leg lymph of normal men. *Scand. J. Lab. Invest.* 37 (1977d) 669
- Olszewski, W.L., A. Engeset: Hemolytic con-

- ment in peripheral lymph of normal men. *Clin. Exp. Immunol.* 32 (1978) 392
- 16 *Olszewski, W.L., A. Engeset*: Intrinsic contractility of leg lymphatics in man. *Lymphology* 12 (1979a) 81
- 17 *Olszewski, W.L., A. Engeset*: Twenty-four hour variation of leg lymph protein of different molecular weight in normal men. *Lymphology. Proc. VIth Int. Cong. Int. Soc. Lymph.* Prague 1977, Ed. Malek, P., et al. Thieme (1979) 98
- 18 *Olszewski, W.L., A. Engeset*: Lymphatic contractility. *New Engl. J. Med.* 300 (1979b) 316
- 19 *Olszewski, W.L., A. Engeset*: Intrinsic contractility of prenodal lymph vessels and lymph flow in human leg. *Amer. J. Physiol.* 239 (1980) H775
- 20 *Olszewski, W.L., Lukomska, B. A. Engeset*: Complement inhibitor in human periopheral lymph. *Arch. Immunol. Ther. Exp.* (1982a) in press
- 21 *Olszewski, W.L., I. Grzelak, A. Engeset*: Cells in lymph draining normal human skin-monoclonal antibody analysis (submitted for publication) 1982b
- 22 *Olszewski, W.L., I. Grzelak, A. Engeset*: High spontaneous and mitogen-induced activity of mononuclear cells in lymph draining normal human skin. *Cl. Exp. Immunol.* (in print) 1982c

Prof. W.L. Olszewski, Dept. of Surg. Res. and Transpl. Medical Research Center, Polish Academy of Sciences, 02004 Warsaw 5 Chalubinskiego, Poland